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## **SEMI-MICRO DETERMINATION OF AMMONIA IN WATER BY THE RUBAZOIC ACID METHOD**

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**A simplified rubazoic acid method for the determination of ammonia on a semi-micro scale is described. The absorbance of a pink-violet aqueous solution of rubazoic acid is measured at 550 nm. The method can be directly applied to fresh waters with NH4'-N concentrations ofbetween 20 and** lo00 **pg. I-'. The semi-micro method was**  compared with the original extraction rubazoic acid method of Procházková (1964) and the results showed good **agreement**  $(R^2 = 0.98, n = 25)$ . The direct nesslerization method gave significant overestimates.

**KEY WORDS: Ammonia determination, rubazoic acid, fresh water.** 

#### INTRODUCTION

The use of bispyrazolone in combination with pyridine and monopyrazolone for the determination of ammonia was first reported by Kruse and Mellon<sup>1</sup> and later applied for sea water by Strickland and Austin<sup>2</sup> and Strickland and Parsons<sup>3</sup>. Procházková<sup>4</sup>, in a detailed study of the reactions involved, eliminated the use of pyridine and defined the individual reaction steps and the final coloured reaction product as rubazoic acid (RA). The specificity of the reaction follows from the reaction mechanism and was experimentally confirmed (neither amines nor amino acids contribute to the final coloured product). Using optimum conditions for the formation of RA, she developed a sensitive extraction method for the determination of ammonia in fresh waters. The procedure by Procházková was further modified for sea water by Johnston<sup>5</sup>. Slawyk and Mac Isaac<sup>6</sup> reported good agreement between the determination of ammonia as indophenol blue and the RA method in non-polluted sea water ( $R^2 = 0.96$ ). Despite the specificity and high sensitivity, the RA method was not widely used, mainly due to its time-consuming extraction steps.

This paper describes a simplified RA method on a semi-micro scale. The proposed method causes a 5-fold reduction in the use of reagents and eliminates the tedious extraction steps and transfer of samples. The absorbance of the pink-violet aqueous solution of RA is measured at **550** nm.

#### MATERIALS AND METHODS

The semi-micro method was performed in test tubes (18 **x** 180 mm) fitted with a ground glass stopper.

The water for sample dilution, standard and reagent solutions, and for glassware rinsing was prepared by redistilling distilled water after the addition of  $KMnO_4$  and  $H_2SO_4$  (RdW). It may be replaced by distilled water passed through a column of a strong cation-exchange resin in the hydrogen form'.

#### *Reagents*

*Citrate-phosphate buffer (pH = 5.8).* Mix 400 ml of 0.1 M citric acid (C<sub>6</sub>H<sub>8</sub>O<sub>7</sub>.H<sub>2</sub>O, 21.01 g.1-l) and 600 ml of 0.2 M Na2HPO4 and stabilize by **0.5 ml** of CHCL. Store at 5°C. Reagent 1.

*Chloramine T(8 g, I').* Dissolve 0.8 g of chloramine T in RdW and dilute to 100 ml. The solution may be stored at 5°C for at least 2 weeks. Reagent 2.

*Bispyrazolone (2 g. l<sup>t</sup>).* Dissolve 0.2 g of 3,3'-dimethyl-5,5'-dioxo-1,1'-diphenyl-(4,4'-bi-2-pyrazoline) in 100 ml of 0.25 M Na<sub>2</sub>CO<sub>3</sub> at 90°C and allow to cool. Prepare daily. Reagent 3.

*Pyrazolone* (2 *g.t').* Dissolve 1 g of **3-methyl-5-oxo-l-phenyl-2-pyrazoline** in 500 ml of hot RdW. The solution is stable for at least 2 months. Reagent 4.

*Standard solution (1 mg.t<sup>1</sup> NH<sub>4</sub><sup>+</sup>-N).* Prepare daily from a 100-fold concentrated solution of NH<sub>4</sub>Cl. Reagent 5.

#### *Procedure*

Add 1 ml of RdW and 1 ml of buffer (reagent 1) to 10 ml of the sample in a test-tube (rinsed by RdW just before analysis) and mix. Add **0.5** ml of chloramine T (reagent **2)** and mix. Allow to stand for 5 min  $\pm$  5 s, add 1 ml of bispyrazolone (reagent 3) and mix. Allow to stand for again **5** min f **5** s, add **2** ml of pyrazolone (reagent 4) and **mix.** Allow to stand for about 20 min, mix the sample again and measure the absorbance at **550** nm in a 2-cm cell against RdW.

At least 2 reaction blanks per each series should be carried through the entire procedure with fiesh RdW.

Standardize the method against 1 ml of the standard solution of NH<sub>4</sub>Cl (reagent 5) added to the another blank (external standard) or another portion of sample (internal standard) instead of 1 ml of RdW and carry them through the whole procedure.

For coloured samples or unfiltered samples containing suspended particles, following procedure is recommended. Take 2 aliquots of the sample, add **0.5** ml of chloramine T to one of them and **0.5** ml of RdW instead of chloramine T to the other one, and continue as described above. Calculate the concentration of ammonia using the corrected absorbance:

where  $A_1$  and  $A_{10}$  are the absorbances of the sample and the blank, respectively, with chloramine T, and  $A_{II}$  and  $A_{II}$  are the absorbances of the sample and the blank, respectively, without chloramine T.

#### RESULTS AND DISCUSSION

The semi-micro method can be directly applied to fresh water samples with concentrations of ammonia ranging from 20 to  $1000 \mu g$ . I 'of N.

The detection limit of the semi-micro method calculated **as** three times of the mean value of the standard deviation (SD) of the blanks was  $18 \mu g$ ,  $1^{-1} NH_4^{\dagger}$ -N. The mean value of the blank was  $26 \pm 6$  µg. 1<sup>-1</sup> NH<sub>4</sub><sup>+</sup>-N ( $\pm$ SD), calculated from 30 duplicates each in different series of samples. The blank value changes from laboratory to laboratory and depends on purity of chemicals, RdW, and atmosphere of the laboratory.

The precision (the average relative standard deviation)<sup>8</sup> was  $1.6\%$  for the concentrations of NH<sub>4</sub><sup>+</sup>-N from 172 to 200 µg.  $1^{-1}$  (n=6) and 7% in the range of 24 to 53 µg. l<sup>-1</sup>NH<sub>4</sub><sup>+</sup>-N (n=4). Each sample was analyzed in *5* parallels.

The accuracy of the semi-micro method was tested by adding a known quantity of ammonia  $(100 \mu g.1^{-1}NH_4^{-1}N)$  to 23 different samples. The average per cent recovery was  $99.3 \pm 5.3 \pm 5.3$ .

The molar absorptivity was determined to be  $10850 \pm 850$  1.mol<sup>-1</sup>.cm<sup>-1</sup> (n=23,  $\pm$ SD) at 550 nm. The sensitivity of the extraction (CHCl<sub>3</sub>) RA method<sup>4</sup> is nearly 2 times higher (the molar absorptivity at 450 mn was  $20700$  1 mol<sup>-1</sup> cm<sup>-1</sup>).

#### *Interferences*

Similarly, as with the extraction method, alkalinity or acidity of the sample greater than 2 mmol.  $1^{-1}$ , temperature of the sample out of the range of  $15 - 20^{\circ}$ C, reducing compounds, and amino acids, amines, CN, and CNS in the molar ratio of  $[N]/[NH<sub>4</sub>]-N]$  greater than 1, interfere with the semi-micro method. Errors can be eliminated by following the guidelines of Procházková<sup>4</sup> or Golterman and Clymo<sup>9</sup>.

The turbidity of the sample caused by particulate substances usually present in some unfiltered samples increases the absorbance. The compensation for this error is described under Procedure. The capability of this step was tested with dispersion of diatomite and clay added to standard solution of  $NH<sub>4</sub>Cl$  (see Table 1).

The colour of the sample increases the absorbance. This error can be compensated for as described under Procedure.

High concentrations of organic matter decrease the recovery of ammonia and cause a negative error. The error can be eliminated by calculating the concentration of ammonia in the sample from the recovery of the internal standard. The capability of the elimination of this error was tested with mixtures of a standard solution of NH<sub>4</sub>Cl with humic acid (n. 2366120, Roth, FRG) and with ligninsulphonic acid (Marasperse N, Marathon, Neenah, USA) (see Table 2).

<b>TURBIDITY</b> <b>NTU</b>	<b>ABSORBANCE</b> 550nm, 2-cm cell	Recovery $ofNH_4^+$ -N	
	$(A_1 - A_{10})^a$	$(AII - AIIO)b$	%
0	100	0	100
8	118	19	99
16	128	29	99
24	153	43	110
32	167	59	108
40	184	80	104

Table **1** Interference of turbidity of dispersion of diatomite and clay added to the standard solution of NH<sub>4</sub>Cl (100 µg. 1<sup>-1</sup> of N) on determination of ammonia.

<sup>a</sup>A<sub>I</sub>, A<sub>IO</sub> is the absorbance of the sample and the blank, respectively; with chloramine T.  $<sup>b</sup>A<sub>II</sub>$ , A<sub>IIO</sub> is the absorbance of the sample and the blank, respectively; without</sup> chloramine T.

The concentration of ammonia was calculated using the absorbance of the sample (after compensation forthe error of its colour) and its internal standard. The obtained results agreed with the real concentrations of ammonia in the mixed samples if the recovery of the internal standard was not less than **50%.** However, at high concentration of organic matter, we recommend to dilute and reanalyse the sample.

Calcium reacts with  $CO<sub>3</sub><sup>2</sup>$  (reagent 3) and turbidity of CaCO<sub>3</sub> appears at concentrations greater than 30 mg. l<sup>-1</sup> of Ca<sup>2+</sup>. Up to 50 mg. l<sup>-1</sup> of Ca<sup>2+</sup> the error of CaCO<sub>3</sub> can be compensated using the same procedure as that used for turbidity. At Ca<sup>2+</sup> concentrations exceeding 50 mg. **I-'** the sample should be diluted.

	Concentration of		Found concentration		Recovery
Organic	<b>DOC</b>	$NH4+-N$		$ofNH_4$ <sup>+</sup> -N	of INT
matter	(mg. I <sup>1</sup> )	$(mg.1-1)$		$(mg. I^{\mathsf{T}})$	(%)
			<b>EXT</b>	<b>INT</b>	
	20	0.10	0.09	0.11	75
	40	0.20	0.14	0.20	70
LA	60	0.30	0.16	0.31	59
	80	0.40	0.17	0.42	45
	100	0.50	0.17	0.43	40
	10	0.10	0.11	0.12	92
	15	0.15	0.13	0.15	83
HA	20	0.20	0.18	0.20	77
	30	0.30	0.21	0.34	67
	50	0.50	0.24	0.51	46

Table **2** Interference of dissolved organic carbon (DOC) added **as** ligninsulphonic acid (LA) or humic acid (HA) *on* determination of ammonia\*.

\*Concentrations of ammonia calculated using the external (EXT) or internal (INT) **standard** 

Sample			$NH_4$ <sup>+</sup> -N $(\mu g/l)$	
		EX	<b>SM</b>	DN
Reservoir	$\mathbf{1}$	47	53	130
	2	29	24	96
	3	178	177	
	4	111	139	207
River water	l	300	335	481
	$\overline{\mathbf{c}}$	214	200	
	3	57	58	74
	$\overline{\mathbf{4}}$	35	38	
	5	470	455	615
	6	320	307	400
	7	446	375	1270
	8	470	490	1240
	9	174	200	435
Fish-pond	ı	124	107	194
	$\boldsymbol{2}$	650	670	1010
	3	350	345	1220
	4	448	489	
Tap water	$\mathbf{I}$	65	58	
	$\overline{\mathbf{c}}$	55	51	
	3	208	199	
	4	$\overline{2}$	3	
Precipitation <sup>a</sup>	$\ldots$	365	340	395
		525	470	645
	$\frac{2}{3}$	146	172	
Waste water <sup>b</sup>	1	226	197	286

Table 3 Comparison of the extraction (EX), proposed semi-micro (SM) and direct nesslerization (DN) methods for water samples **(n=5).** 

<sup>a</sup>After 10-fold dilution.

<sup>b</sup>After 50-fold dilution.

#### *Comparison of semi-micro, extraction and direct nesslerization methods*

The comparison was performed on 25 samples of water of various types (Table 3).

The results demonstrate that the semi-micro and the extraction RA methods concurred with each other  $(R^2 = 0.98, n = 24$ ; paired t-test showed no statistically significant difference at the *5%* level) whilst the direct nesslerization (DN) method' gave significant overestimates. The average ratio of the results obtained by the DN and RA methods was  $1.88 \pm 0.77$  (n = 16,  $\pm$ SD). The average percent NH<sub>4</sub><sup>+</sup>-N recovery of the DN method was 127  $\pm$  39 (n = 16,  $\pm$ SD).

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